

Formal Total Synthesis of Neocarzinostatin Chromophore

Shoji Kobayashi,*,^{†,‡} Makiko Hori,[†] Guang Xing Wang,[†] and Masahiro Hirama*,[†]

Departments of Chemistry, Graduate Schools of Science, Tohoku University, Sendai 980-8578, and Osaka Prefecture University, Sakai 599-8531, Japan

koba@c.s.osakafu-u.ac.jp

Received September 28, 2005



An efficient route to the neocarzinostatin chromophore aglycon has been developed. The present strategy involves a stereoselective intramolecular acetylide—aldehyde cyclization to form the C5–C6 bond, followed by efficient installation of α -epoxide, naphthoate, and carbonate functionalities. The C8–C9-olefin was introduced by using the Martin sulfurane dehydration reaction to furnish the highly reactive aglycon.

Introduction

Neocarzinostatin (NCS), the first enediyne antibiotic,¹ was isolated from a culture of *Streptomyces carzinostaticus* var. F-41 in 1965.² Its potent antibacterial and antitumor activities derive from the inhibition of DNA synthesis and DNA degradation in cells.³ The poly(styrene-maleic acid)–NCS conjugate, namely, SMANCS, shows promising clinical activity against primary hepatocellular carcinoma.^{3c,4} NCS is composed of a very unstable chromophore⁵ and a carrier apoprotein.⁶ The bioactive small molecular chromophore (NCS-chr, **1**) consists of a

(1) Reviews on enediyne antibiotics: (a) Nicolaou, K. C.; Dai, W.-M. Angew. Chem., Int. Ed. Engl. **1991**, 30, 1387–1416. (b) Grissom, J. W.; Gunawardena, G. U.; Klingberg, D.; Huang, D. Tetrahedron **1996**, 52, 6453–6518. (c) Xi, Z.; Goldberg, I. H. DNA-damaging Enediyne Compounds. In Comprehensive Natural Products Chemistry; Barton, S. D., Nakanishi, K., Meth-Cohn, O., Eds.; Pergamon: Oxford, 1999; Vol. 7, pp 553–592.

(2) Ishida, N.; Mizugaki, K.; Kumagai, K.; Rikimaru, M. J. Antibiot. 1965, 18, 68.

(3) (a) Ono, Y.; Yatanabe, Y.; Ishida, N. Biochim. Biophys. Acta **1966**, 119, 46–58. (b) Beerman, T. A.; Goldberg, I. H. Biochem. Biophys. Res. Commun. **1974**, 59, 1254–1261. (c) Neocarzinostatin, The Past, Present, and Future of an Anticancer Drug, Maeda, H., Edo, K., Ishida, N., Eds.; Springer: Tokyo, 1997.

(4) (a) Maeda, H. Adv. Drug Delivery Rev. **1991**, 6, 181–202. (b) Maeda, H. Adv. Drug Delivery Rev. **2001**, 46, 169–185.

(5) Isolation of the chromophore: (a) Napier, M. A.; Holmquist, B.; Strydom, D. J.; Goldberg, I. H. *Biochem. Biophys. Res. Commun.* **1979**, 89, 635–642. (b) Koide, Y.; Ishii, F.; Hasuda, K.; Koyama, Y.; Edo, K.; Katamine, S.; Kitame, F.; Ishida, N. *J. Antibiot.* **1980**, *33*, 342. naphthoate intercalator, a 2'-methylamino sugar, and an epoxy bicyclo[7,3,0]dodecadienediyne core (Figure 1).⁷ The naked chromophore is highly sensitive to light,⁸ heat,⁹ base,¹⁰ and nucleophiles¹¹ and readily undergoes cycloaromatization to generate bioactive diradical species.^{1c,12} This chemical instability, complex structure, and fascinating mode of action make **1** a challenging target for total synthesis.

(8) (a) Uesawa, Y.; Kuwahara, J.; Sugiura, Y. *Biochem. Biophys. Res. Commun.* **1989**, *164*, 903–911. (b) Gomibuchi, T.; Fujiwara, K.; Nehira, T.; Hirama, M. *Tetrahedron Lett.* **1993**, *36*, 5753–5756. (c) Gomibuchi, T.; Hirama, M. J. Antibiot. **1995**, *48*, 738–740.

(9) Edo, K.; Akiyama-Murai, Y.; Saito, K.; Mizugaki, M.; Koide, Y.; Ishida, N. J. Antibiot. **1988**, 41, 1272.

(10) (a) Hensens, O. D.; Helms, G. L.; Zink, D. L.; Chin, D.-H.; Kappen,
L. S.; Goldberg, I. H. *J. Am. Chem. Soc.* **1993**, *115*, 11030–11031. (b)
Stassinopoulos, A.; Ji, J.; Gao, X.; Gordberg, I. H. *Science* **1996**, *272*, 1943–1946.

(11) (a) Myers, A. G. *Tetrahedron Lett.* **1987**, 28, 4493–4496. (b) Tanaka, T.; Fujiwara, K.; Hirama, M. *Tetrahedron Lett.* **1990**, 31, 5947–5950. (c) Fujiwara, K.; Sakai, H.; Tanaka, T.; Hirama, M. *Chem. Lett.* **1994**, 457. (d) Sugiyama, H.; Yamashita, K.; Fujiwara, T.; Saito, I. *Tetrahedron* **1994**, 50, 1311–1325. (e) Myers, A. G.; Arvedson, S. P.; Lee, R. W. J. Am. Chem. Soc. **1996**, 118, 4725–4726.

(12) (a) Goldberg, I. H. Acc. Chem. Res. **1991**, 24, 191–198. (b) Sugiyama, H.; Fujiwara, T.; Kawabata, H.; Yoda, N.; Hirayama, N.; Saito, I. J. Am. Chem. Soc. **1992**, 114, 5573–5578 and references therein.

10.1021/jo052031o CCC: \$33.50 © 2006 American Chemical Society Published on Web 12/20/2005

^{*} To whom correspondence should be addressed. Phone: +81-72-254-9698. Fax: +81-72-254-9698.

[†] Tohoku University.

[‡] Osaka Prefecture University.

^{(6) (}a) Crystal structure of NCS: Kim, K.-H.; Kwon, B.-M.; Myers, A. G.; Rees, D. C. *Science* **1993**, *262*, 1042–1046. (b) Solution structure of NCS: Tanaka, T.; Hirama, M.; Fujita, K.; Imajo, S.; Ishiguro, M. J. Chem. Soc., Chem. Commun. **1993**, 1205–1207.

^{(7) (}a) Chromophore structure: Edo, K.; Mizugaki, M.; Koide, Y.; Seto, H.; Furihata, K.; Otake, N.; Ishida, N. *Tetrahedron Lett.* **1985**, *26*, 331–334. (b) Carbohydrate stereochemistry: Edo, K.; Akiyama, Y.; Saito, K.; Mizugaki, M.; Koide, Y.; Ishida, N. *J. Antibiot.* **1986**, *39*, 1615. (c) Chromophore stereochemistry: Myers, A. G.; Proteau, P. J.; Handel, T. M. *J. Am. Chem. Soc.* **1988**, *110*, 7212–7214.





FIGURE 1. Structure of the neocarzinostatin chromophore and the strategy for the synthesis of the aglycon.

Although many notable synthetic approaches in this area have been reported,¹³ total synthesis of the nine-membered enediyne natural product has been elusive.^{14,15} With respect to the NCSchr (1), only one successful total synthesis has been achieved by Myers and co-workers.¹⁶ Their synthesis involves the stereocontrolled glycosidic coupling between the 2'-methylamino sugar and the labile aglycon 2 in unprotected form.^{16b,c} This highly practical methodology avoids the need for protecting the 2'-methylamino group and minimizes synthetic manipulations after installation of the labile enediyne functionality. Furthermore, aglycon 2 serves as a viable substrate for glycosylation; the fucosyl analogue and the radiolabeled NCS-chr can both be prepared by introducing the suitable glycoside into 2.^{16c,17} We therefore considered aglycon 2 a valuable synthetic target for further developing NCS-chr (1). Herein we report a synthesis of 2, featuring convergent assembly of the functionalized cyclopentenone with the acetylenic fragment, followed by stereoselective intramolecular acetylide–aldehyde cyclization^{18,19} between the C5 and C6 positions^{14b,20} (Figure 1). The crucial C8–C9-olefin was efficiently installed in the final stage by dehydration of the C9-tertiary alcohol.

Results and Discussion

Nine-Membered Ring Closure. The synthesis began with the optically active (1R, 4S, 5S)-4,5-bis(methoxymethoxy)-2cyclopenten-1-ol (3), which was readily prepared on a large scale from commercially available methyl D-glucopyranoside by recently developed RCM technology^{21,22} (Scheme 1). Before further details of the synthesis are presented, it should be emphasized that the highly oxygenated nature of several intermediates required an elaborate protecting group strategy. Thus, the hydroxyl group of 3 was protected as the acid-stable TBDPS ether, and the MOM ethers were cleaved selectively in the presence of a catalytic amount of ZrCl₄.²³ Other deprotection conditions, including the use of alkylboron bromide or PPTS in t-BuOH, gave disappointing results. Selective oxidation of the allylic alcohol with MnO_2 afforded the α -hydroxy enone 4. Despite its moderate sensitivity under oxidative conditions, the MPM ether was determined to be the most appropriate protecting group of the C10-alcohol moiety due to its robustness to basic, nucleophilic, and mildly acidic conditions. However, because of the chemical instability of enone 4 under both acidic and basic conditions, it was necessary to employ 4-methoxybenzyl 2,2,2-trichloroacetimidate²⁴ at low temperature to install the MPM group. Subsequent α -iodination²⁵ afforded the Sonogashira coupling precursor 5.

Parallel synthetic studies had suggested that slightly harsh conditions were required to remove the acetonide group of the

(19) For leading examples of intramolecular acetylide-aldehyde cyclizations to construct 10-membered enediynes, see: (a) Kende, A. S.; Smith, C. A. *Tetrahedron Lett.* **1988**, *29*, 4217–4220. (b) Cabal, M. P.; Coleman, R. S.; Danishefsky, S. J. *J. Am. Chem. Soc.* **1990**, *112*, 3253–3255.

(20) (a) Sato, I.; Toyama, K.; Kikuchi, T.; Hirama, M. Synlett **1998**, 1308–1310. (b) Inoue, M.; Hatano, S.; Kodama, M.; Sasaki, T.; Kikuchi, T.; Hirama, M. Org. Lett. **2004**, 6, 3833–3836. (c) Inoue, M.; Sasaki, T.; Hatano, S.; Hirama, M. Angew. Chem., Int. Ed. **2004**, 43, 6500–6505. (d) Inoue, M.; Kikuchi, T.; Hirama, M. Tetrahedron Lett. **2004**, 45, 6439–6442.

(21) Koyama, Y.; Lear, M. J.; Yoshimura, F.; Ohashi, I.; Mashimo, T.; Hirama, M. Org. Lett. **2005**, *7*, 267–270.

(22) For our previous synthesis of chiral trihydroxylated cyclopentenes akin to **3**, see: (a) Toyama, K.; Iguchi, S.; Oishi, T.; Hirama, M. *Synlett* **1995**, 1243–1244. (b) Toyama, K.; Iguchi, S.; Sakazaki, H.; Oishi, T.; Hirama, M. *Bull. Chem. Soc. Jpn.* **2001**, *74*, 997–1008.

(23) Sharma, G. V. M.; Reddy, K. L.; Lakshmi, P. S.; Krishna, P. R. *Tetrahedron Lett.* **2004**, *45*, 9229–9232.

(24) Audia, J. E.; Boisvert, L.; Patten, A. D.; Villalobos, A.; Danishefsky, S. J. J. Org. Chem. **1989**, *54*, 3738–3740.

(25) Johnson, C. R.; Adams, J. P.; Braun, M. P.; Senanayake, C. B. W.; Wovkulich, P. M.; Uskokovic, M. R. *Tetrahedron Lett.* **1992**, *33*, 917–918.

⁽¹³⁾ For construction of the nine-membered diyne systems, see: (a) Wender, P. A.; Harmata, M.; Jeffrey, D.; Mukai, C.; Suffert, J. *Tetrahedron Lett.* **1988**, *29*, 909–912. (b) Wender, P. A.; McKinney, J. A.; Mukai, C. J. *Am. Chem. Soc.* **1990**, *112*, 5369–5370. (c) Doi, T.; Takahashi, T. J. Org. Chem. **1991**, *56*, 3465–3467. (d) Tanaka, H.; Yamada, H.; Matsuda, A.; Takahashi, T. Synlett **1997**, *4*, 381–383. (e) Magnus, P.; Carter, R.; Davies, M.; Elliott, J.; Pitterna, T. Tetrahedron **1996**, *52*, 6283–6306. (f) Myers, A. G.; Harrington, P. M.; Kuo, E. Y. J. Am. Chem. Soc. **1991**, *113*, 694–695. (g) Myers, A. G.; Goldberg, S. D. Angew. Chem., Int. Ed. **2000**, *39*, 2732–2735.

^{(14) (}a) Total synthesis of N1999-A2: Kobayashi, S.; Reddy, R. S.; Sugiura, Y.; Sasaki, D.; Miyagawa, N.; Hirama, M. J. Am. Chem. Soc. 2001, 123, 2887–2888. (b) Kobayashi, S.; Ashizawa, S.; Takahashi, Y.; Sugiura, Y.; Nagaoka, M.; Lear, M. J.; Hirama, M. J. Am. Chem. Soc. 2001, 123, 11294–11295. (c) Synthesis of the kedarcidin chromophore aglycon: Myers, A. G.; Hogan, P. C.; Hurd, A. R.; Goldberg, S. D. Angew. Chem., Int. Ed. 2002, 41, 1062–1067.

⁽¹⁵⁾ Total syntheses of 10-membered enediyne antibiotics have been reported. (a) Calicheamicin γ_1^{I} : Nicolaou, K. C.; Hummel, C. W.; Nakada, M.; Shibayama, K.; Pitsinos, E. N.; Saimoto, H.; Mizuno, Y.; Baldenius, K.-U.; Smith, A. L. J. Am. Chem. Soc. **1993**, 115, 7625–7635. (b) Hitchcock, S. A.; Chu-Moyer, M. Y.; Boyer, S. H.; Olson, S. H.; Danishefsky, S. J. J. Am. Chem. Soc. **1995**, 117, 5750–5756. (c) Dynemicin A: Shair, M. D.; Yoon, T.-Y.; Mosny, K. K.; Chou, T. C.; Danishefsky, S. J. J. Am. Chem. Soc. **1996**, 118, 9509–9525. (d) Myers, A. G.; Tom, N. J.; Fraley, M. E.; Cohen, S. B.; Madar, D. J. J. Am. Chem. Soc. **1997**, 119, 6072–6094.

^{(16) (}a) Myers, A. G.; Hammond, M.; Wu, Y.; Xiang, J.-N.; Harrington, P. M.; Kuo, E. Y. J. Am. Chem. Soc. 1996, 118, 10006-10007. (b) Myers, A. G.; Liang, J.; Hammond, M.; Harrington, P. M.; Wu, Y.; Kuo, E. Y. J. Am. Chem. Soc. 1998, 120, 5319-5320. (c) Myers, A. G.; Glatthar, R.; Hammond, M.; Harrington, P. M.; Kuo, E. Y.; Liang, J.; Schaus, S. E.; Wu, Y.; Xiang, J.-N. J. Am. Chem. Soc. 2002, 124, 5380-5401.

⁽¹⁷⁾ Myers, A. G.; Liang, J.; Hammond, M. Tetrahedron Lett. **1999**, 40, 5129–5133.

^{(18) (}a) Iida, K.; Hirama, M. J. Am. Chem. Soc. **1994**, *116*, 10310–10311. (b) Sato, I.; Akahori, Y.; Iida, K.; Hirama, M. Tetrahedron Lett. **1996**, *37*, 5135–5138. (c) Kawata, S.; Yoshimura, F.; Irie, J.; Ehara, H.; Hirama, M. Synlett **1997**, 250–252. (d) Das, P.; Mita, T.; Lear, M. J.; Hirama, M. Chem. Commun. **2002**, 2624–2625. (e) Mita, T.; Kawata, S.; Hirama, M. Chem. Lett. **1998**, 959–960. (f) Myers, A. G.; Harrington, P. M.; Kuo, E. Y. J. Am. Chem. Soc. **1991**, *113*, 694–695. (g) Myers, A. G.; Harrington, P. M.; Kwon, B.-M. J. Am. Chem. Soc. **1992**, *114*, 1086–1087.

SCHEME 1. Construction of the Nine-Membered Ring



C13,C14-diol in the right half of the molecule.^{14a,b} Heeswijk et al. showed that cyclopentylidene ketals of 1,2-diols tend to be 3 times more susceptible to acid-catalyzed hydrolysis than their corresponding acetonides.²⁶ Therefore, the C13,C14-diol moiety of the acetylenic fragment 6^{27} was protected as its cyclopentylidene ketal prior to coupling. Palladium-catalyzed coupling²⁸ between 5 and 6, followed by TES protection, afforded the expected envne 7 in high yield. Addition of propargylmagnesium bromide to 7 proceeded quantitatively, but in a nonstereoselective manner. During this transformation, partial loss of pivaloyl ester was observed. Therefore, the crude products were treated with PivCl without purification to yield a separable mixture of 9R- and 9S-isomers (8 and 9) in a 1.2:1 ratio. Interestingly, the proportion of the 9R-isomer (8) in the present case was less than that reported previously,^{20a} and there was no significant change in product ratio upon addition of ZnCl₂.²⁹ It was thought that the protecting groups on the cyclopentenone moiety played an important role in defining the selectivity of the Grignard addition. Since NOE data were inconclusive, the stereochemistry of the C9-tertiary alcohol of 9 was determined by derivatization to its MP-acetal 10 upon anhydrous DDQ treatment.

Sequential protecting group manipulation and oxidation³⁰ of **8** provided the key cyclization precursor **12**. The 9*R*-configured aldehyde **12** underwent ring closure smoothly using the CeCl₃/LiN(TMS)₂-mediated cyclization protocol³¹ to afford the C4,C5-

(30) Dess, D. B.; Martin, J. C. J. Org. Chem. 1983, 48, 4155-4156.

(31) Imamoto, T.; Takiyama, N.; Nakamura, K.; Hatajima, T.; Kamiya, Y. J. Am. Chem. Soc. 1989, 111, 4392–4398. *trans*-alcohol **14** as a single stereoisomer in 73% yield. The remarkable *trans*-selectivity observed in **14** was consistent with a number of previous results^{14b,18a-d,20a-c} and presumably arose via the energetically favored transition state **13**, in which the cerium-chelated carbonyl oxygen is sterically remote from the neighboring TES group. Furthermore, it should be pointed out that the corresponding 9*S*-isomer derived from **9** was inactive to the nine-membered ring closure, another strong indication that the relative stereochemistry of the precursors is responsible for efficient cyclization.^{14a,b,18a,b}

Synthesis of the Neocarzinostatin Chromophore Aglycon. On the basis of previous research, the C4,C5-epoxide was strategically constructed by taking advantage of the different reactivities of the three TES groups in 14 under fluoridemediated deprotection conditions (Scheme 2).14b Treatment of 14 with MsCl provided the corresponding mesylate, which was exposed to 2 equiv of TBAF at -35 °C to afford the desired epoxide 16 as a major product. The minor diol intermediate 15 was readily converted into 16 by potassium carbonate in EtOH. Importantly, the C9-O(TES) group was left untouched during these selective deprotection and tandem epoxidation steps, thereby allowing the manipulations which followed. Alcohol 16 thus obtained was condensed with naphthoic acid $(17)^{32}$ without concern for bisnaphthoate formation.^{14b} Further investigation revealed that the C9-hydroxyl group must be protected during the next carbonate formation step; otherwise the desired carbonate was formed in less than 35% yield. To overcome this inefficiency, selective conditions to cleave the cyclopentylidene ketal in the presence of the acid-labile TES and epoxide functionalities were investigated. It was found that the cyclopentylidene group could be removed selectively upon treatment with dilute aqueous hydrofluoric acid in CH₃CN to give the desired triol 18 in 69% yield. Facile carbonate formation could

⁽²⁶⁾ van Heeswijk, W. A. R.; Goedhart, J. B.; Vliegenthart, J. F. G. Carbohydr. Res. 1977, 58, 337–344.

^{(27) (}a) Nakatani, K.; Arai, K.; Hirayama, N.; Matsuda F.; Terashima, S. *Tetrahedron* **1992**, *48*, 633–650. (b) Kobayashi, S.; Das, P.; Wang, G.

X.; Mita, T.; Lear, M. J.; Hirama, M. *Chem. Lett.* **2002**, 300–301. (28) Sonogashira, K.; Tohda, Y.; Hagihara, N. *Tetrahedron Lett.* **1975**,

<sup>50, 4467–4470.
(29)</sup> Tanaka, H.; Yamada, H.; Matsuda, A.; Takahashi, T. Synlett 1997, 381–383.

^{(32) (}a) Takahashi, K.; Suzuki, T.; Hirama, M. *Tetrahedron Lett.* **1992**, *33*, 4603–4604. (b) Takahashi, K.; Tanaka, T.; Suzuki, T.; Hirama, M. *Tetrahedron* **1994**, *50*, 1327–1340.





then be realized, forming 20 after removal of the C9-O(TES) group (66% yield in two steps). In view of the instability of aglycon 2, it was advisable at this stage to cleave the MPM ether.^{14a,b} The MPM group in **20** was replaced by a TES group via DDQ oxidation and selective protection. In contrast to the behavior of the C10-deoxy substrate, which underwent dehydration at 0 °C,14b alcohol 21 remained unaltered upon treatment with a large excess of Ms₂O, Et₃N, and DMAP; only incomplete formation of the C9-mesylate resulted. Addition of DBU to the resulting mesylate led to rapid decomposition, while other dehydration conditions including Tf₂O and 2,6-lutidine^{14a} were not effective. The steric hindrance due to the C10-O(TES) and the C11-naphthoate groups is likely to inhibit such dehydrations. After numerous trials, the desired enediyne 22 was finally obtained by using the Martin sulfurane dehydrating reagent.³³ Remarkably, the reaction proceeded quantitatively as determined by NMR within 5 min at room temperature. The protected aglycon 22 was found to be stable to silica gel but rapidly decomposed in neat form. The final deprotection was carefully achieved with a system of TFA-THF-H₂O (1:10:5) to furnish the highly unstable aglycon 2. TLC monitoring of the reaction indicated that clean deprotection occurred, but the purified aglycon 2 was extremely sensitive to concentration and degraded much more rapidly than the protected aglycon 22, both in solution and, especially, in neat form. It seems likely that the bulkiness of the protecting groups again contributed to the stability of the chromophore core.³⁴ All spectroscopic data of synthetic 2 were identical to those reported in the literature.^{16a} In addition, the high-resolution mass spectrum (ESI-FT-ICR-MS (m/z) [M + Na]⁺, calcd for C₂₈H₂₀NaO₉ 523.1000, found

523.1002) was obtained. An efficient conversion of aglycon 2 to NCS-chr (1) has previously been accomplished by Myers' group.^{16b,c}

Conclusion

An efficient route to the neocarzinostatin chromophore aglycon was developed. The present synthesis involves convergent assembly of the functionalized cyclopentenone with the acetylenic fragment, followed by stereocontrolled nine-membered ring closure. After introduction of epoxide, naphthoate, and carbonate functionalities, a central dienediyne system was elaborated by using sulfurane-mediated dehydration reaction. A highly efficient protecting group strategy was also developed to access the sensitive substrate.

Experimental Section

Enone 4. To a solution of allyl alcohol 3 (11.0 g, 54.1 mmol) in DMF (54 mL) were added imidazole (9.21 g, 135 mmol) and (TBDPS)Cl (15.2 mL, 59.5 mmol). The mixture was stirred for 6 h at room temperature and poured into water. The resulting mixture was extracted with hexane $(2\times)$, and the combined organic layer was washed with brine and dried over anhydrous Na₂SO₄. Concentration of the solution gave the corresponding TBDPS ether (19.9 g), which was subjected to the next reaction without purification: ¹H NMR (500 MHz, CDCl₃) δ 1.09 (s, 9H), 3.39 (s, 3H), 3.42 (s, 3H), 4.30 (t, 1H, J = 4.3 Hz), 4.37 (br s, 1H), 4.69 (br s, 1H), 4.73 (d, 1H, J = 7.0 Hz), 4.75 (d, 1H, J = 7.0 Hz), 4.75 (d, 1H, J = 7.0Hz), 4.81 (d, 1H, J = 7.0 Hz), 5.58 (d, 1H, J = 6.5 Hz), 5.76 (d, 1H, J = 6.5 Hz), 7.38–7.45 (m, 6H), 7.71–7.77 (m, 4H); ¹³C NMR (125 MHz, CDCl₃) δ 19.5, 27.2, 55.7, 56.0, 81.0, 85.1, 91.9, 96.5 (2C), 127.9, 128.0, 130.0, 130.1, 131.9, 133.8, 134.5, 135.1, 136.2, 136.2; FT-IR (neat) v 3017, 2932, 2891, 2858, 2823, 1962, 1894, 1827, 1589, 1471, 1428, 1369, 1307, 1261, 1212 cm⁻¹.

To a solution of the above TBDPS ether (19.9 g) in *i*PrOH (98 mL) was added $ZrCl_4$ (1.14 g, 4.89 mmol) at 90 °C. The mixture

^{(33) (}a) Arhart, R. J.; Martin, J. C. *J. Am. Chem. Soc.* **1982**, *104*, 1116–1118. (b) Myers et al. also succeeded in introducing the C12–C1-olefin using the Martin sulfurane: see ref 16.

⁽³⁴⁾ Myers, A. G.; Kort, M. E.; Hammond, M. J. Am. Chem. Soc. 1997, 119, 2965–2972.

was stirred for 4.5 h at 90 °C and cooled to 0 °C. The reaction mixture was filtered through a pad of Celite, and the filtrate was concentrated. The residue was purified by column chromatography (Florisil, hexane–hexane/AcOEt = 1) to give the corresponding diol (12.5 g, 35.2 mmol, 65% in two steps) as a colorless oil: $[\alpha]_D^{30.0} - 9.0^{\circ}$ (*c* 0.442, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.08 (s, 9H), 4.05 (br s, 1H), 4.35 (br s, 1H), 4.52 (br s, 1H), 5.70 (br d, 1H, *J* = 6.5 Hz), 5.73 (br d, 1H, *J* = 6.5 Hz), 7.39–7.45 (m, 6H), 7.67–7.75 (m, 4H); ¹³C NMR (125 MHz, CDCl₃) δ 19.3, 27.0, 80.2, 82.0, 89.8, 128.0, 128.1, 130.1, 130.2, 133.0, 133.7, 134.1, 134.5, 135.9; FT-IR (neat) ν 3362, 3071, 2931, 2858, 1471, 1427, 1362 cm⁻¹; MALDI-TOF-MS (*m*/*z*) [M + Na]⁺, calcd for C₂₁H₂₆NaO₃Si 377.1549, found 377.1529.

To a solution of the above diol (7.60 g, 21.4 mmol) in CH₂Cl₂ (428 mL) was added MnO₂ (27.9 g, 321 mol). The mixture was stirred for 7 h at room temperature and filtered. The filter cake was washed with hexanes–EtOAc, and the filtrate was concentrated. The residue was purified by flash column chromatography (silica gel, hexane/AcOEt = 3.4) to give enone **4** (5.07 g, 14.4 mmol, 67%) as a pale yellow oil: $[\alpha]_D^{30.0}$ +6.6° (*c* 2.36, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.12 (s, 9H), 2.46 (br s, 1H), 4.29 (br s, 1H), 4.76 (br d, 1H, *J* = 1.5 Hz), 6.15 (dd, 1H, *J* = 6.5, 1.0 Hz), 7.16 (dd, 1H, *J* = 6.5, 1.8 Hz), 7.36–7.49 (m, 6H), 7.71–7.76 (m, 4H); ¹³C NMR (125 MHz, CDCl₃) δ 19.3, 26.9, 27.0, 78.1, 81.7, 127.7, 127.8, 127.9, 127.9, 128.0, 130.3, 130.3, 131.5, 132.8, 133.4, 135.8, 135.9, 136.0, 161.0, 203.9; FT-IR (neat) ν 3436, 3071, 2932, 2892, 2858, 1729, 1472, 1427, 1363 cm⁻¹; HR-ESI-FT-ICR-MS (*m*/*z*) [M + Na]⁺, calcd for C₂₁H₂₄NaO₃Si 375.1387, found 375.1387.

Iodide 5. To a solution of enone 4 (4.09 g, 11.6 mmol) in CH₂-Cl₂ (116 mL) were added 4-methoxybenzyl 2,2,2-trichloroacetimidate (4.92 g, 17.4 mmol) and BF3•Et2O (147 µL, 1.16 µmol) at -65 °C. The mixture was stirred for 1.5 h at -65 °C and quenched with saturated NaHCO₃ solution. The resulting mixture was extracted with AcOEt $(2\times)$, and the combined organic layer was washed with brine and dried over anhydrous MgSO₄. After concentration, the residue was purified by flash column chromatography (silica gel, hexane/AcOEt = 12 to 10 to 8 to 5 to 3 to 2) to give the corresponding MPM ether (2.81 g, 5.95 mmol, 51%) as a yellow oil: $[\alpha]_D^{29.0} - 7.9^\circ$ (c 1.07, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.09 (s, 9H), 3.81 (s, 3H), 4.16 (d, 1H, J = 3.0Hz), 4.58 (d, 1H, J = 11 Hz), 4.89 (br s, 1H), 4.90 (d, 1H, J = 11Hz), 6.06 (br d, 1H, J = 6.5 Hz), 6.86 (d, 2H, J = 6.0 Hz), 7.05 (dd, 1H, J = 6.0, 2.0 Hz), 7.24 (dd, 2H, J = 6.5 Hz), 7.36–7.46 (m, 6H), 7.66–7.73 (m, 4H); ¹³C NMR (125 MHz, CDCl₃) δ 19.3, 27.0, 55.4, 71.6, 72.7, 77.0, 86.0, 113.9, 113.9, 128.0, 129.6, 129.8, 129.9, 130.2, 132.7, 132.8, 133.5, 135.9, 136.0, 159.5, 160.0, 203.5; FT-IR (neat) v 3071, 2999, 2932, 2858, 1963, 1888, 1727, 1613, 1587, 1514, 1463, 1428, 1391, 1361, 1327, 1302, 1248 cm⁻¹; MALDI-TOF-MS (m/z) [M + Na]⁺, calcd for C₂₉H₃₂NaO₄Si 495.1968, found 495.1990.

To a solution of the above MPM ether (3.02 g, 6.36 mmol) in THF (64 mL) were added pyridine (1.54 mL, 19.1 mmol) and I₂ (3.23 g, 12.7 mmol). The mixture was stirred for 30 min at room temperature, diluted with Et2O, and quenched with saturated Na₂S₂O₃ solution. The resulting mixture was extracted with AcOEt $(2\times)$, and the combined organic phase was washed with brine and dried over anhydrous MgSO₄. After concentration, the residue was purified by flash column chromatography (silica gel, hexane/ AcOEt = 20) to give iodide 5 (3.09 g, 5.17 mmol, 81%) as a colorless oil: $[\alpha]_D^{29.0} - 61.0^{\circ}$ (*c* 0.602, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.08 (s, 9H), 3.81 (s, 3H), 4.17 (d, 1H, J = 2.5 Hz), 4.58 (d, 1H, J = 11 Hz), 4.84 (t, 1H, J = 2.5 Hz), 4.92 (d, 1H, J = 11Hz), 6.86 (d, 2H, J = 8.0 Hz), 7.23 (d, 2H, J = 9.0 Hz), 7.37-7.40 (m, 4H), 7.45 (d, 1H, J = 2.5 Hz), 7.45-7.48 (m, 2H), 7.65-7.70 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 19.3, 26.9, 55.4, 72.7, 78.4, 83.4, 102.3, 113.9, 128.1, 128.1, 129.4, 130.1, 130.4, 130.4, 132.5, 133.0, 135.9, 135.9, 159.6, 165.7, 198.4; FT-IR (neat) v 3071, 2932, 2858, 1730, 1613, 1587, 1514, 1470, 1427, 1392, 1352, 1302, 1248 cm⁻¹; HR-ESI-FT-ICR-MS (m/z) [M + Na]⁺, calcd for C₂₉H₃₁NaIO₄Si 621.0929, found 621.0930.

Cyclopentylidene Ketal 6. To a solution of (2S)-2-hydroxy-2-[(4*R*)-2,2-dimethyl-1,3-dioxolan-4-yl]-3-butyn-1-yl 2,2-dimethylpropanoate²⁷ (0.219 g, 0.810 mmol) in MeOH (8.1 mL) was added TsOH·H₂O (15.4 mg, 81.0 μ mol). The mixture was stirred for 4 h at room temperature and concentrated. The residue was dissolved in MeOH (8.1 mL) and stirred for another 5 h at room temperature. After concentration, 1,1-dimethoxycyclopentane (213 µL, 1.62 mmol) and cyclopentanone (3.5 mL) were added, and the mixture was stirred for 30 min at room temperature. The reaction mixture was quenched with Et₃N (500 μ L) followed by saturated NaHCO₃ solution and extracted with AcOEt $(2\times)$. The combined organic layer was washed with brine and dried over anhydrous MgSO₄. After concentration, the remaining cyclopentanone was distilled out azeotropically with toluene. The residue was purified by flash column chromatography (silica gel, hexane/AcOEt = 8 to 6 to 5) to give cyclopentylidene ketal 6, which was further purified by recrystallization from hexane to afford 6 (0.138 g, 0.510 mmol, 63%) as colorless needles: mp 102–103 °C (hexane); $[\alpha]_D^{28.0}$ +21.7° (c 1.00, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.25 (s, 9H), 1.67–1.70 (m, 4H), 1.74–1.77 (m, 2H), 1.87–1.95 (m, 2H), 2.49 (s, 1H), 3.03 (s, 1H), 4.05 (dd, 1H, J = 7.3, 5.8 Hz), 4.12 (t, 1H, J = 5.3 Hz), 4.14 (dd, 1H, J = 7.3, 5.3 Hz), 4.19 (d, 1H, J = 12 Hz), 4.40 (d, 1H, J = 12 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 23.4, 23.9, 27.3, 36.1, 36.4, 39.1, 65.9, 68.0, 71.7, 74.9, 77.6, 81.4, 120.5, 179.1; FT-IR (KBr) v 3443, 3270, 2945, 1708, 1669, 1541, 1475 cm⁻¹. Anal. Calcd for C₁₆H₂₄O₅: C, 64.84; H, 8.16. Found: C, 64.83; H, 8.28.

Enone 7. A mixture of iodide 5 (3.09 g, 5.17 mmol), acetylenic fragment 6 (1.84 g, 6.20 mmol), and CuI (0.394 g, 2.07 mmol) was dissolved in DMF (50 mL). To the solution were added *i*Pr₂-NEt (2.70 mL, 15.5 mmol) and (Ph₃P)₄Pd (0.598 g, 0.517 mmol), and the mixture was stirred for 1 h at room temperature. The reaction mixture was diluted with hexane-AcOEt and quenched with saturated NH₄Cl solution. The resulting mixture was extracted with AcOEt $(2\times)$, and the combined organic layer was washed with brine and dried over anhydrous MgSO₄. After concentration, the residue was purified by flash column chromatography (silica gel, hexane/AcOEt = 5 to 4 to 3 to 2 to 1) to give the coupling product (3.86 g, 5.03 mmol, 97%) as a yellow oil: $[\alpha]_D^{27.0} - 21.0^{\circ}$ (c 1.00, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.07 (s, 9H), 1.21 (s, 9H) 1.61-1.70 (m, 4H), 1.72-1.77 (m, 2H), 1.78-1.90 (m, 2H), 3.39 (s, 1H), 3.81 (s, 3H), 4.04 (dd, 1H, J = 11, 9.0 Hz), 4.09–4.13 (m, 2H), 4.16 (d, 1H, J = 2.5 Hz), 4.18 (d, 1H, J = 11 Hz), 4.40 (d, 1H, J = 11 Hz), 4.56 (d, 1H, J = 11 Hz), 4.84 (t, 1H, J = 2.5 Hz), 4.89 (d, 1H, J = 11 Hz), 6.85 (d, 2H, J = 8.5 Hz), 7.04 (d, 1H, J = 2.5 Hz), 7.22 (d, 2H, J = 8.5 Hz), 7.35–7.39 (m, 4H), 7.43-7.46 (m, 2H), 7.64-7.69 (m, 4H); FT-IR (neat) v 3437, 3072, 2960, 1731, 1613, 1588, 1514, 1471, 1428, 1395, 1337, 1283, 1249 cm⁻¹; MALDI-TOF-MS (m/z) [M + Na]⁺, calcd for C₄₅H₅₄NaO₉-Si 789.3435, found 789.3447.

To a solution of the above coupling product (1.98 g, 2.58 mmol) in CH₂Cl₂ (26 mL) were added 2,6-lutidine (1.80 mL, 15.5 mmol) and (TES)OTf (1.16 mL, 5.16 mmol) at -70 °C. The mixture was stirred for 40 min at -70 °C and quenched with saturated NaHCO₃ solution. The resulting mixture was extracted with hexane $(2\times)$, and the combined organic layer was washed with brine and dried over anhydrous Na₂SO₄. After concentration, the residue was purified by flash column chromatography (silica gel, hexanehexane/AcOEt = 25 to 20) to give TES ether 7 (2.27 g, 2.58 mmol, 100%) as a colorless oil: $[\alpha]_D^{27.0} - 31.9^\circ$ (c 0.66, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.67 (q, 6H, J = 8.0 Hz), 0.92 (t, 9H, J = 8.0 Hz), 1.08 (s, 9H), 1.21 (s, 9H) 1.60-1.68 (m, 4H), 1.71-1.74 (m, 2H), 1.76–1.79 (m, 1H), 1.81–1.87 (m, 1H), 3.81 (s, 3H), 4.01 (dd, 1H, J = 8.3, 6.8 Hz), 4.05 (dd, 1H, J = 8.3, 5.8 Hz), 4.10 (d, 1H, J = 11 Hz), 4.14 (t, 1H, J = 6.3 Hz), 4.18 (d, 1H, J = 3.0 Hz), 4.31 (d, 1H, J = 11 Hz), 4.59 (d, 1H, J = 11 Hz), 4.86 (t, 1H, J = 2.5 Hz), 4.92 (d, 1H, J = 11 Hz), 6.86 (d, 2H, J

= 8.5 Hz), 7.00 (d, 1H, J = 2.5 Hz), 7.23 (d, 2H, J = 8.5 Hz), 7.36–7.39 (m, 4H), 7.44–7.47 (m, 2H), 7.65–7.70 (m, 4H); ¹³C NMR (125 MHz, CDCl₃) δ 6.0, 7.1, 15.4, 19.3, 23.5, 23.8, 26.9, 27.3, 36.5, 36.6, 39.0, 53.6, 55.5, 66.0, 66.5, 67.6, 72.8, 73.0, 75.6, 77.6, 77.9, 85.7, 95.1, 113.9, 120.4, 127.7, 128.0, 128.1, 129.6, 130.0, 130.3, 130.4, 132.6, 133.2, 135.9, 136.0, 159.6, 160.2, 198.5; FT-IR (neat) ν 3072. 2957, 2876, 1732, 1613, 1588, 1514, 1463, 1428, 1395, 1336, 1283, 1249 cm⁻¹; MALDI-TOF-MS (m/z) [M + Na]⁺, calcd for C₅₁H₆₈NaO₉Si₂ 903.4300, found 903.4302.

Alcohols 8 and 9. To a suspension of magnesium turnings (1.46 g, 60.0 mmol) and mercury(II) chloride (5 mg) in Et₂O (30 mL) was slowly added a solution of propargyl bromide (3.56 mL, 40.0 mmol) in Et₂O (10 mL) over 20 min at room temperature. The mixture was refluxed for 1.5 h to give a 1.0 M solution of propargylmagnesium bromide. To a solution of enone 7 (2.27 g, 2.58 mmol) in toluene (86 mL) was added the above propargylmagnesium bromide solution (5.16 mL, 5.16 mmol) at -78 °C. The mixture was stirred for 15 min at -78 °C and quenched with saturated NH₄Cl solution. The resulting mixture was extracted with hexanes-EtOAc $(2\times)$, and the combined organic layer was washed with brine and dried over anhydrous Na2SO4. After concentration, the residue was dissolved in pyridine (10 mL) and treated with PivCl (396 μ L, 3.22 mmol) and DMAP (~5 mg). The mixture was stirred for 17 h and concentrated. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc = 20 to 10) to give a less polar (9R)-alcohol, 8 (1.24 g, 1.34 mmol, 52%), and a polar (9S)-alcohol, 9 (1.05 g, 1.14 mmol, 44%), as pale yellow oils. Data for 8: $[\alpha]_D^{26.0} - 28.9^\circ$ (c 2.00, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.65 (q, 6H, J = 7.8 Hz), 0.91 (t, 9H, J = 7.8Hz), 1.06 (s, 9H), 1.23 (s, 9H) 1.65-1.71 (m, 6H), 1.76 (t, 1H, J = 2.3 Hz), 1.90 (m, 2H), 2.47 (dd, 1H, J = 16, 2.8 Hz), 2.59 (dd, 1H, J = 16, 2.8 Hz), 3.34 (s, 1H), 3.81 (s, 3H), 4.00 (t, 1H)J = 6.5 Hz), 4.11 - 4.17 (m, 2H), 4.13 (d, 1H, J = 11 Hz), 4.14 (d, 1H, J = 5.5 Hz), 4.32 (d, 1H, J = 11 Hz), 4.56 (d, 1H, J = 11Hz), 4.78 (d, 1H, J = 11 Hz), 4.82 (dd, 1H, J = 5.5, 1.5 Hz), 5.76 (d, 1H, J = 2.0 Hz), 6.85 (d, 2H, J = 8.5 Hz), 7.24 (d, 2H, J = 8.5Hz), 7.30-7.43 (m, 6H), 7.65-7.70 (m, 4H); ¹³C NMR (125 MHz, $CDCl_3$) δ 6.0, 7.0, 19.3, 23.3, 23.8, 26.4, 27.1, 27.3, 35.9, 36.0, 39.1, 55.4, 66.3, 67.4, 70.8, 72.9, 73.2, 77.4, 78.5, 79.6, 80.7, 83.2, 92.2, 93.9, 113.7, 114.0, 120.4, 127.7, 127.8, 127.9, 128.0, 128.1, 129.4, 129.7, 129.8, 129.9, 130.7, 133.4, 134.0, 135.9, 136.0, 136.0, 139.3, 159.2, 178.0; FT-IR (neat) v 3443, 3311, 2958, 1732, 1613, 1588, 1514, 1463, 1428, 1336, 1248, 1112 cm⁻¹; MALDI-TOF-MS (m/z) [M + Na]⁺, calcd for C₅₄H₇₂NaO₉Si₂ 943.4613, found 943.4591. Data for **9**: [α]_D^{26.0} –11.4° (*c* 1.77, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.68 (q, 6H, J = 7.7 Hz), 0.93 (t, 9H, J = 7.8 Hz), 1.09 (s, 9H), 1.23 (s, 9H) 1.58-1.69 (m, 4H), 1.72-1.77 (m, 2H), 1.79-1.91 (m, 2H), 2.07 (t, 1H, J = 2.5 Hz), 2.66 (dd, 1H, J = 17, 2.8 Hz), 2.76 (dd, 1H, J = 17, 2.5 Hz), 3.21 (s, 1H), 3.80 (s, 3H), 4.02 (dd, 1H, J = 8.0, 7.0 Hz), 4.07 (dd, 1H, J = 8.5, 6.5 Hz), 4.12 (d, 1H, J = 2.5 Hz), 4.14 (d, 1H, J = 11 Hz), 4.16 (t, 1H, J = 6.5 Hz), 4.34 (d, 1H, J = 12 Hz), 4.43 (d, 1H, J = 11Hz), 4.52 (d, 1H, J = 11 Hz), 4.81 (t, 1H, J = 2.5 Hz), 5.82 (d, 1H, J = 2.0 Hz), 6.85 (d, 2H, J = 8.0 Hz), 7.16 (d, 2H, J = 8.5Hz), 7.38-7.41 (m, 4H), 7.44-7.47 (m, 2H), 7.65-7.72 (m, 4H); ¹³C NMR (125 MHz, CDCl₃) δ 6.0, 7.0, 19.3, 23.5, 23.8, 27.0, 27.3, 29.0, 36.3, 36.5, 39.0, 55.4, 66.5, 67.7, 70.9, 73.0, 73.1, 77.7, 80.2, 80.3, 81.4, 81.7, 86.6, 92.2, 113.9, 120.2, 127.9, 128.0, 129.7, 129.7, 130.1, 130.1, 130.7, 133.3, 133.6, 135.9, 136.0, 139.6, 159.6, 178.0; FT-IR (neat) v 3522, 3311, 2957, 1732, 1613, 1588, 1514, 1462, 1428, 1337, 1249, 1111 cm⁻¹; MALDI-TOF-MS (*m/z*) $[M + Na]^+$, calcd for C₅₄H₇₂NaO₉Si₂ 943.4613, found 943.4599.

Identification of the C9 Stereochemistry. To a solution of alcohol 9 (30.0 mg, 0.0326 mmol) in CH_2Cl_2 (8 mL) was added DDQ (29.6 mg, 0.130 mmol). The mixture was stirred for 19 h at room temperature and concentrated. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc = 12 to 11 to 10 to 6 to 4) to give MP-acetal **10** (1.1:1 diastereomeric mixture, 12.9 mg, 0.0140 mmol, 43%) as a colorless syrup: ¹H

NMR (500 MHz, CDCl₃) δ 0.64 (q, 6H, J = 8.0 Hz), 0.65 (q, 6H, J = 8.0 Hz), 0.87 (t, 9H, J = 8.0 Hz), 0.88 (t, 9H, J = 8.0 Hz), 1.06 (s, 9H), 1.07 (s, 9H), 1.20 (s, 9H), 1.21 (s, 9H), 1.53-1.66 (m, 8H), 1.68-1.73 (m, 4H), 1.75-1.88 (m, 4H), 2.10 (t, 1H, J =2.5 Hz), 2.10 (t, 1H, J = 2.5 Hz), 2.74 (dd, 1H, J = 17, 3.0 Hz), 2.87 (dd, 1H, J = 17, 2.5 Hz), 3.05 (dd, 1H, J = 17, 3.0 Hz), 3.10 (dd, 1H, J = 17, 2.5 Hz), 3.76 (s, 3H), 3.80 (s, 3H), 3.99 (t, 1H, J = 6.5 Hz), 4.01 (t, 1H, J = 6.5 Hz), 4.06 (t, 1H, J = 6.5 Hz), 4.07 (t, 1H, J = 6.5 Hz), 4.12 (d, 1H, J = 11 Hz), 4.13 (t, 1H, J = 6.5 Hz), 4.14 (d, 1H, J = 11 Hz), 4.14 (t, 1H, J = 6.5 Hz), 4.32 (d, 1H, J = 11 Hz), 4.33 (d, 1H, J = 11 Hz), 4.66 (br s, 1H), 4.75 (br d, 1H, J = 2.5 Hz), 4.75 (br s, 1H), 4.77 (d, 1H, J = 2.5Hz), 5.42 (s, 1H), 5.71 (dd, 1H, J = 2.5, 1.0 Hz), 6.00 (dd, 1H, J = 2.5, 1.0 Hz), 6.07 (s, 1H), 6.79 (d, 2H, J = 9.0 Hz), 6.86 (d, 2H, J = 8.0 Hz), 7.25 (d, 2H, J = 9.0 Hz), 7.35–7.47 (m, 14H), 7.62-7.69 (m, 8H); ESI-TOF-MS (m/z) [M + Na]⁺, calcd for C₅₄H₇₀NaO₉Si₂ 941.4456, found 941.4456.

TES Ether 11. To a solution of alcohol 8 (944 mg, 1.03 mmol) in THF (10 mL) was added TBAF (1.0 M solution in THF, 2.56 mL, 2.56 mmol). The mixture was stirred for 1.5 h at room temperature and concentrated. The residue was purified by flash column chromatography (silica gel, hexane/AcOEt = 2 to 1) to give the corresponding triol (429 mg, 0.755 mmol, 74%) as a colorless amorphous solid: $[\alpha]_D^{27.0} - 6.9^\circ$ (*c* 2.35, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.25 (s, 9H), 1.63–1.76 (m, 6H), 1.92–1.96 (m, 2H), 2.04 (s, 1H), 2.64 (t, 2H, J = 2.8 Hz), 3.18 (s, 1H), 3.58 (s, 1H), 3.80 (s, 3H), 3.89 (d, 1H, J = 5.0 Hz), 4.04 (dd, 1H, J = 9.0, 7.0 Hz), 4.09 (dd, 1H, J = 7.0, 3.3 Hz), 4.16 (d, 1H, J = 12 Hz), 4.31 (dd, 1H, J = 8.5, 3.3 Hz), 4.52 (d, 1H, J = 11 Hz), 4.64 (d, 1H, J = 12 Hz), 4.68 (dd, 1H, J = 5.0, 1.5 Hz), 4.83 (d, 1H, J = 11 Hz), 6.04 (d, 1H, J = 2.0 Hz), 6.89 (d, 2H, J = 9.0 Hz), 7.34 (d, 2H, J = 8.5 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 23.3, 23 8, 26.1, 27.3, 36.0, 36.0, 39.1, 55.4, 66.1, 67.7, 71.4, 72.2, 72.9, 77.1, 77.4, 79.5, 79.9, 83.5, 92.5, 93.9, 114.0, 120.7, 129.0, 129.5, 130.4, 138.2, 159.3, 179.2; FT-IR (film) v 3417, 3307, 2961, 2875, 1730, 1613, 1514, 1480, 1398, 1337, 1285, 1248, 1172, 1107 cm⁻¹; MALDI-TOF-MS (m/z) [M + Na]⁺, calcd for C₃₂H₄₀NaO₉ 591.2570, found 591.2592.

To a solution of the above triol (429 mg, 0.755 mmol) in CH₂-Cl₂ (10 mL) were added 2,6-lutidine (703 µL, 6.04 mmol) and (TES)OTf (680 μ L, 3.02 mmol) at -50 °C. The mixture was stirred for 1 h at -50 °C and quenched with saturated NaHCO₃ solution. The resulting mixture was extracted with hexanes-EtOAc $(2\times)$, and the combined organic layer was dried over anhydrous Na₂-SO₄. After concentration, the residue was purified by flash column chromatography (silica gel, hexane-hexane/AcOEt = 10) to give tris(TES ether) 11 (619 mg, 0.680 mmol, 90%) as a pale yellow syrup: [α]_D^{22.0} –23.9° (*c* 2.45, CHCl₃); ¹H NMR (500 MHz, $CDCl_3$) δ 0.61 (q, 6H, J = 8.0 Hz), 0.63 (q, 6H, J = 8.0 Hz), 0.71 (q, 6H, J = 8.0 Hz), 0.93 (t, 9H, J = 8.0 Hz), 0.95 (t, 9H, J = 8.0Hz), 0.97 (t, 9H, J = 8.0 Hz), 1.23 (s, 9H), 1.60–1.69 (m, 4H), 1.72–1.75 (m, 2H), 1.83–1.94 (m, 2H), 1.87 (t, 1H, J = 2.5 Hz), 2.50 (dd, 1H, J = 16, 2.8 Hz), 2.73 (dd, 1H, J = 16, 2.3 Hz), 3.81 (s, 3H), 4.01 (d, 1H, J = 4.5 Hz), 4.02 (dd, 1H, J = 8.5, 7.0 Hz), 4.07 (dd, 1H, J = 8.5, 7.0 Hz), 4.12 (d, 1H, J = 12 Hz), 4.17 (t, 1H, J = 6.8 Hz), 4.35 (d, 1H, J = 11 Hz), 4.63 (d, 1H, J = 12Hz), 4.77 (d, 1H, J = 11 Hz), 4.83 (dd, 1H, J = 5.0, 2.0 Hz), 6.04 (d, 1H, J = 2.0 Hz), 6.88 (dd, 2H, J = 7.0, 2.0 Hz), 7.32 (d, 2H, J = 8.0 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 5.2, 6.0, 6.1, 6.5, 6.6, 6.9, 7.1, 7.2, 23.5, 23.8, 27.0, 27.3, 36.4, 36.6, 39.0, 55.4, 66.5, 67.7, 69.9, 72.2, 73.0, 77.6, 78.7, 80.6, 82.6, 86.2, 90.9, 95.0, 113.7, 113.7, 120.1, 129.0, 129.4, 129.7, 130.9, 139.8, 159.1, 177.9; FT-IR (neat) v 3313, 2955, 1732, 1614, 1587, 1515, 1463, 1416, 1397, 1336, 1282, 1247 cm⁻¹; MALDI-TOF-MS (m/z) [M + Na]⁺, calcd for C₅₀H₈₂NaO₉Si₃ 933.5164, found 933.5162.

Aldehyde 12. To a solution of pivalate 11 (688 mg, 0.755 mmol) in Et₂O (10 mL) was added DIBAL (0.94 M solution in hexane, 3.21 mL, 3.02 mmol) at -78 °C. The mixture was stirred for 30 min at -78 °C and quenched with saturated Rochelle salt solution.

The resulting mixture was stirred at room temperature until the two phases were clearly separated. The organic phase was separated, and the aqueous phase was extracted with hexane. The combined organic layer was dried over anhydrous Na₂SO₄. After concentration, the residue was purified by flash column chromatography (silica gel, hexane/AcOEt = 7) to give the corresponding alcohol (553 mg, 0.672 mmol, 89%) as a colorless oil: $[\alpha]_D^{20.0} - 28.9^{\circ}$ (*c* 0.59, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.61 (q, 6H, J = 7.5Hz), 0.63 (q, 6H, J = 7.5 Hz), 0.74 (q, 6H, J = 7.5 Hz), 0.93 (t, 9H, J = 7.5 Hz), 0.97 (t, 18H, J = 7.5 Hz), 1.60–1.69 (m, 4H), 1.70-1.81 (m, 2H), 1.85-1.95 (m, 2H), 1.88 (t, 1H, J = 2.5 Hz), 2.18 (dd, 1H, J = 10, 3.8 Hz), 2.51 (dd, 1H, J = 16, 2.5 Hz), 2.71 (dd, 1H, J = 16, 2.5 Hz), 3.68 (dd, 1H, J = 11, 3.3 Hz), 3.77 (t, 1H, J = 10 Hz), 3.81 (s, 3H), 4.00 (d, 1H, J = 5.0 Hz), 4.03 (dd, 1H, J = 8.5, 7.0 Hz), 4.07 (dd, 1H, J = 8.5, 6.0 Hz), 4.17 (t, 1H, J = 6.5 Hz), 4.62 (d, 1H, J = 12 Hz), 4.77 (d, 1H, J = 11 Hz), 4.83 (dd, 1H, J = 5.0, 2.0 Hz), 6.02 (d, 1H, J = 2.0 Hz), 6.89 (d, 2H, J = 9.0 Hz), 7.32 (d, 2H, J = 8.5 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 5.2, 6.1, 6.4, 6.9, 7.1, 7.2, 23.5, 23.7, 27.0, 36.3, 36.7, 55.4, 55.4, 66.6, 68.1, 70.0, 72.2, 74.4, 77.9, 78.7, 78.8, 80.6, 83.0, 86.1, 91.4, 95.0, 113.8, 120.2, 129.0, 129.5, 130.9, 139.5, 159.2; FT-IR (neat) v 3499, 3312, 2955, 1614, 1514, 1462, 1415, 1336, 1247, 1110 cm⁻¹; MALDI-TOF-MS (m/z) [M + Na]⁺, calcd for C₄₅H₇₄NaO₈Si₃ 849.4589, found 849.4596.

To a solution of the above alcohol (482 mg, 0.583 mmol) in CH_2Cl_2 (12 mL) were added pyridine (236 μ L, 2.92 mmol) and Dess-Martin periodinane (494 mg, 1.17 mmol). The mixture was stirred for 2 h at room temperature, diluted with hexane, and quenched with saturated Na₂S₂O₃ solution and then saturated NaHCO₃ solution. The resulting mixture was extracted with hexane $(2\times)$, and the combined organic layer was dried over anhydrous MgSO₄. After concentration, the residue was purified by column chromatography (Florisil, hexane/AcOEt = 10) to give aldehyde **12** (433 mg, 0.525 mmol, 90%) as a colorless syrup: $[\alpha]_D^{24.0} + 2.1^\circ$ $(c \ 0.570, \text{CHCl}_3)$; ¹H NMR (500 MHz, CDCl₃) $\delta \ 0.60$ (q, 6H, J =7.8 Hz), 0.63 (q, 6H, J = 7.8 Hz), 0.71 (q, 6H, J = 7.8 Hz), 0.92 (t, 9H, J = 7.8 Hz), 0.97 (t, 9H, J = 7.8 Hz), 0.97 (t, 9H, J = 7.8Hz), 1.63–1.69 (m, 4H), 1.71–1.76 (m, 2H), 1.84–1.95 (m, 2H), 1.88 (brs, 1H), 2.52 (dd, 1H, J = 16, 1.8 Hz), 2.73 (dd, 1H, J =16, 1.5 Hz), 3.81 (s, 3H), 4.01 (d, 1H, J = 5.0 Hz), 4.04 (dd, 1H, J = 8.5, 7.0 Hz), 4.12 (dd, 1H, J = 8.5, 6.0 Hz), 4.16 (t, 1H, J =6.5 Hz), 4.62 (d, 1H, J = 12 Hz), 4.77 (d, 1H, J = 11 Hz), 4.83 (d, 1H, J = 5.0 Hz), 6.08 (s, 1H), 6.89 (d, 2H, J = 8.0 Hz), 7.32 (d, 2H, J = 8.0 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 5.2, 6.1, 6.4, 6.9, 7.0, 7.2, 23.5, 23.8, 27.0, 36.2, 36.4, 55.4, 55.4, 66.3, 70.0, 70.1, 72.3, 78.1, 78.7, 78.7, 79.7, 80.5, 85.1, 86.1, 87.9, 95.0, 113.8, 120.9, 129.0, 129.1, 130.8, 140.5, 159.2, 195.5, 195.6; FT-IR (neat) v 3312, 2956, 2877, 1745, 1614, 1515, 1462, 1415, 1337, 1248, 1172, 1120 cm⁻¹; MALDI-TOF-MS (m/z) [M + Na]⁺, calcd for C₄₅H₇₂NaO₈Si₃ 847.4433, found 847.4418.

Alcohol 14. To well-ground anhydrous CeCl₃ (1.00 g, 4.06 mmol) was slowly added THF (50 mL) with vigorous stirring at 0 °C. The suspension was stirred for 36 h at room temperature. A cooled (0 °C) solution of LiN(TMS)₂ in THF, which was freshly prepared from 1,1,1,3,3,3-hexamethyldisilazane (1.04 mL, 4.91 mmol) and *n*BuLi (1.56 M solution in hexane, 2.85 mL, 4.45 mmol) in THF (50 mL), was added to the above suspension at -40 °C. The mixture was gradually warmed to -20 °C over 1 h, and a solution of aldehyde 12 (386 mg, 0.468 mmol) in THF (50 mL) was added in one portion. The resulting mixture was allowed to warm to room temperature and stirred for 1 h. The reaction mixture was quenched with phosphate buffer (pH 7.0) and filtered through a pad of Celite. The precipitate was washed with Et₂O, and the combined filtrate was concentrated. The residue was again extracted with hexane, and the organic extracts were washed with saturated NH₄Cl solution and brine and dried over anhydrous Na₂SO₄. After concentration, the residue was purified by flash column chromatography (silica gel, hexane/EtOAc = 10 to 8) to give the ninemembered alcohol 14 (282 mg, 0.342 mmol, 73%) as a yellow

syrup: [α]_D^{24.0} -46.4° (*c* 0.450, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.64 (q, 6H, J = 7.5 Hz), 0.67 (q, 6H, J = 8.3 Hz), 0.73 (q, 6H, J = 7.8 Hz), 0.96 (t, 9H, J = 7.5 Hz), 0.97 (t, 9H, J = 8.0Hz), 0.98 (t, 9H, J = 7.5 Hz), 1.57–1.67 (m, 4H), 1.70–1.77 (m, 2H), 1.79-1.85 (m, 2H), 2.25 (dd, 1H, J = 17, 3.8 Hz), 2.36 (br s, 1H), 2.67 (dd, 1H, J = 17, 1.8 Hz), 3.81 (s, 3H), 4.01 (d, 1H, J = 4.5 Hz), 4.06 (dd, 1H, J = 8.0, 7.0 Hz), 4.09 (dd, 1H, J = 8.5, 6.0 Hz), 4.35 (t, 1H, J = 6.5 Hz), 4.54 (dd, 1H, J = 4.8, 2.3 Hz), 4.58 (d, 1H, *J* = 11 Hz), 4.67 (br s, 1H), 4.70 (d, 1H, *J* = 11 Hz), 5.87 (d, 1H, J = 2.0 Hz), 6.89 (d, 2H, J = 9.0 Hz), 7.26 (d, 2H, J = 8.5 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 5.2, 5.9, 6.5, 7.0, 23.4, 23.8, 29.5, 29.9, 36.3, 36.6, 55.5, 67.0, 72.7, 75.3, 75.4, 78.7, 78.8, 78.9, 82.8, 84.5, 85.3, 91.0, 91.8, 95.3, 97.2, 113.9, 119.6, 129.3, 130.3, 131.4, 135.8, 135.9, 159.4; FT-IR (neat) v 3469, 2955, 2877, 1614, 1515, 1457, 1415, 1337, 1302, 1247, 1108 cm⁻¹; HR-ESI-FT-ICR-MS (m/z) [M + Na]⁺, calcd for C₄₅H₇₂NaO₈Si₃ 847.4427, found 847.4425.

Epoxide 16. To a solution of the nine-membered ring alcohol 14 (983 mg, 1.19 mmol) in CH₂Cl₂ (24 mL) were added Et₃N (1.66 mL, 11.9 mmol) and MsCl (138 μ L, 1.79 mmol) at -20 °C. The mixture was stirred for 15 min at -20 °C and guenched with saturated NaHCO₃ solution. The resulting mixture was extracted with hexane $(2\times)$, and the combined organic layer was washed with brine and dried over anhydrous MgSO₄. Concentration of the solution gave the corresponding mesylate (1.06 g), which was subjected to the next reaction without purification: ¹H NMR (500 MHz, CDCl₃) δ 0.65 (q, 6H, J = 8.0 Hz), 0.67 (q, 6H, J = 8.0Hz), 0.77 (q, 6H, J = 8.0 Hz), 0.98 (t, 9H, J = 8.0 Hz), 0.98 (t, 18H, J = 8.0 Hz), 1.59-1.70 (m, 4H), 1.71-1.77 (m, 2H), 1.79-1.85 (m, 2H), 2.29 (dd, 1H, J = 17, 4.0 Hz), 2.72 (dd, 1H, J = 17, 1.8 Hz), 3.12 (s, 3H), 3.81 (s, 3H), 3.40 (dd, 1H, *J* = 9.0, 7.0 Hz), 4.03 (d, 1H, J = 4.5 Hz), 4.04 (dd, 1H, J = 9.0, 6.0 Hz), 4.32 (dd, 1H, J = 7.0, 6.0 Hz), 4.55 (dd, 1H, J = 4.5, 2.0 Hz), 4.57 (d, 1H, J = 11 Hz), 4.70 (d, 1H, J = 11 Hz), 5.35 (dd, 1H, J = 4.0, 1.8 Hz), 5.93 (d, 1H, J = 2.0 Hz), 6.89 (d, 2H, J = 8.5 Hz), 7.26 (d, 2H, J = 9.0 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 5.2, 5.8, 6.5, 6.9, 7.0, 7.2, 23.4, 23.9, 29.7, 29.9, 36.1, 36.4, 38.9, 55.5, 66.6, 72.7, 78.4, 78.8, 80.1, 80.8, 81.3, 84.4, 91.6, 95.0, 95.9, 97.3, 114.0, 119.6, 128.5, 129.4, 130.1, 130.8, 136.6, 159.5; FT-IR (neat) v 2956, 2877, 1614, 1515, 1462, 1415, 1370, 1336, 1248, 1180, 1109 cm⁻¹; HR-ESI-FT-ICR-MS (m/z) [M + Na]⁺, calcd for C₄₆H₇₄NaO₁₀SSi₃ 925.4203, found 925.4207.

To a solution of the above mesylate (1.06 g) in THF (23 mL) was added TBAF (1.0 M solution in THF, 2.57 mL, 2.57 mmol) at -40 °C. The mixture was stirred for 15 min at -40 to -35 °C and quenched with saturated NH₄Cl solution. The resulting mixture was extracted with AcOEt $(3\times)$, and the combined organic layer was washed with brine and dried over anhydrous MgSO4. After concentration, the residue was purified by flash column chromatography (silica gel, hexane/AcOEt = 8 to 3 to 2 to 1) to give epoxide 16 (393 mg, 0.679 mmol, 58% in two steps) and diol 15 (121 mg, 0.180 mmol, 15% in two steps) as yellow syrups. To a solution of diol 15 (121 mg, 0.180 mmol) in EtOH (3.6 mL) was added K₂CO₃ (24.9 mg, 0.180 mmol). The mixture was stirred for 17 h at room temperature and concentrated. The residue was purified by flash column chromatography (silica gel, hexane/AcOEt = 3) to give epoxide 16 (54.2 mg, 0.0936 mmol, 52% from 15): $[\alpha]_D{}^{23.0}$ $+7.6^{\circ}$ (c 0.810, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.64 (q, 6H, J = 8.0 Hz), 0.95 (t, 9H, J = 8.0 Hz), 1.66–1.73 (m, 4H), 1.74-1.81 (m, 2H), 1.88-1.97 (m, 2H), 2.35 (d, 1H, J = 18 Hz), 2.74 (d, 1H, J = 18 Hz), 3.52 (s, 1H), 3.81 (s, 3H), 3.94 (d, 1H, J = 5.0 Hz), 4.01 (dd, 1H, J = 5.5, 8.0 Hz), 4.05 (t, 1H, J = 5.8Hz), 4.13 (dd, 1H, J = 6.0, 8.0 Hz), 4.53–4.56 (m, 1H), 4.65 (d, 1H, J = 11.5 Hz), 4.69 (d, 1H, J = 11.5 Hz), 5.91 (d, 1H, J = 2.0 Hz), 6.90 (d, 2H, J = 8.0 Hz), 7.29 (d, 2H, J = 8.0 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 6.4, 7.2, 23.5, 23.8, 29.3, 36.2, 36.3, 54.3, 55.5, 64.3, 67.5, 72.9, 74.4, 78.1, 83.2, 85.1, 88.1, 88.6, 95.1, 96.3, 114.1, 120.8, 129.4, 130.2, 131.7, 134.3, 159.6; FT-IR (neat) v 3480, 2954, 2875, 1613, 1514, 1455, 1337, 1302, 1247, 1108 cm⁻¹; HR-

ESI-FT-ICR-MS (m/z) [M + Na]⁺, calcd for C₃₃H₄₂NaO₇Si 601.2592, found 601.2592.

Triol 18. To a suspension of epoxide 16 (86.8 mg, 0.150 mmol) and naphthoic acid 17 (105 mg, 0.450 mmol) in CH₂Cl₂ (3.0 mL) were added EDC·HCl (115 mg, 0.600 mmol) and DMAP (1.8 mg, 0.015 mmol) at 0 °C. The mixture was stirred for 10 min at 0 °C, and naphthoic acid 17 (21.5 mg, 0.0926 mmol), EDC·HCl (17.8 mg, 0.0926 mmol), and DMAP (1.1 mg, 0.0093 mmol) were further added. The resulting mixture was stirred for 10 min at 0 °C and quenched with water. The mixture was extracted with EtOAc $(3 \times)$, and the combined organic layer was washed with brine and dried over anhydrous Na₂SO₄. After concentration, the residue was purified by flash column chromatography (silica gel, hexane/ AcOEt = 8 to 6) to give the corresponding ester (74.3 mg, 0.0937) mmol, 64%) as a colorless syrup: $[\alpha]_D^{24.0} - 14.5^{\circ}$ (*c* 0.510, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.64 (q, 6H, J = 7.8 Hz), 0.93 (t, 9H, J = 7.8 Hz), 1.66–1.73 (m, 4H), 1.74–1.80 (m, 2H), 1.88– 1.98 (m, 2H), 2.52 (d, 1H, J = 18 Hz), 2.65 (s, 3H), 2.82 (d, 1H, J = 18 Hz), 3.56 (s, 1H), 3.72 (s, 3H), 3.83 (s, 3H), 4.03 (dd, 1H, J = 7.5, 5.0 Hz), 4.06 (t, 1H, J = 5.3 Hz), 4.14 (dd, 1H, J = 7.5,5.8 Hz), 4.40 (d, 1H, J = 5.0 Hz), 4.61 (d, 1H, J = 11 Hz), 4.65 (d, 1H, J = 11 Hz), 5.93 (dd, 1H, J = 5.0, 2.0 Hz), 6.12 (d, 1H, J = 2.0 Hz), 6.72 (d, 2H, J = 8.5 Hz), 6.92 (br s, 1H), 7.05 (d, 1H, J = 9.5 Hz), 7.16 (d, 2H, J = 8.5 Hz), 8.07 (br s, 1H), 8.07 (d, 1H, J = 9.5 Hz), 12.12 (s, 1H); FT-IR (neat) ν 2954, 2876, 1644, 1614, 1514, 1455, 1414, 1376, 1337, 1306, 1248, 1203 cm⁻¹; HR-ESI-FT-ICR-MS (m/z) [M + Na]⁺, calcd for C₄₆H₅₂NaO₁₀Si 815.3222, found 815.3226.

To a solution of the above ester (147 mg, 0.186 mmol) in CH₃-CN (6.2 mL) was added 46% aqueous hydrofluoric acid (67.4 μ L) at 0 °C. The mixture was stirred for 1.6 h at 0 °C and quenched with saturated NaHCO3 solution. The resulting mixture was diluted with Et2O and stirred for 30 min at 0 °C. The mixture was extracted with AcOEt $(3\times)$, and the combined organic layer was washed with brine and dried over anhydrous MgSO₄. After concentration, the residue was purified by flash column chromatography (silica gel, hexane/AcOEt = 2 to 1 to 0.33, containing 2.5% MeOH) to give triol 18 (93.0 mg, 0.128 mmol, 69%) as a colorless syrup and tetrol 19 (31.8 mg, 0.0520 mmol, 28%) as colorless solid. Data for 18: $[\alpha]_D^{22.0}$ -55.1° (*c* 0.482, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.93 (br s, 1H), 2.38 (br s, 1H), 2.63 (s, 3H), 2.68 (d, 1H, J = 18Hz), 2.86 (d, 1H, J = 18 Hz), 3.65 (s, 3H), 3.72 (s, 3H), 3.76 (s, 1H), 3.80 (br dd, 1H, J = 12, 5.7 Hz), 3.96 (br dd, 1H, J = 12, 2.5 Hz), 4.02 (br dd, 1H, J = 4.5, 3.5 Hz), 4.42 (d, 1H, J = 5.0 Hz), 4.63 (d, 1H, J = 12 Hz), 4.71 (d, 1H, J = 12 Hz), 6.00 (dd, 1H, J = 5.0, 2.0 Hz), 6.12 (d, 1H, J = 2.0 Hz), 6.66 (d, 2H, J = 8.5Hz), 6.88 (d, 1H, J = 2.0 Hz), 7.04 (d, 1H, J = 9.5 Hz), 7.16 (d, 2H, J = 8.5 Hz), 7.91 (d, 1H, J = 2.0 Hz), 8.06 (d, 1H, J = 9.5Hz), 12.02 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 6.4, 7.2, 20.3, 29.5, 53.0, 55.3, 55.3, 63.5, 64.0, 69.6, 73.0, 80.8, 83.4, 85.0, 87.9, 88.5, 91.1, 97.3, 104.2, 105.0, 114.0, 115.8, 116.3, 123. 2, 129.3, 129.4, 130.6, 133.2, 134.2, 134.3, 137.3, 159.5, 159.6, 165.2, 172.0; FT-IR (film) v 3417, 2954, 2876, 1644, 1615, 1514, 1463, 1412, 1377, 1345, 1306, 1248, 1204 cm⁻¹; HR-ESI-FT-ICR-MS (m/z) $[M + Na]^+$ calcd for $C_{41}H_{46}NaO_{10}Si$ 749.2752, found 749.2755. Data for **19**: $[\alpha]_D^{22.0} - 110^\circ$ (*c* 0.610, CHCl₃); ¹H NMR (500 MHz, $CDCl_3$) δ 0.64 (q, 6H, J = 8.0 Hz), 0.94 (t, 9H, J = 8.0 Hz), 1.98 (br s, 1H), 2.36 (br s, 1H), 2.54 (d, 1H, J = 18 Hz), 2.65 (s, 3H), 2.82 (d, 1H, J = 18 Hz), 3.70 (s, 1H), 3.72 (s, 3H), 3.79 (br dd, 1H, J = 12, 5.5 Hz), 3.83 (s, 3H), 3.96 (br dd, 1H, J = 12, 2.5 Hz), 4.04-4.08 (m, 1H), 4.40 (d, 1H, J = 5.0 Hz), 4.61 (d, 1H, J = 12 Hz), 4.65 (d, 1H, J = 12 Hz), 5.93 (dd, 1H, J = 5.0, 2.0Hz), 6.13 (d, 1H, J = 2.0 Hz), 6.73 (d, 2H, J = 8.5 Hz), 6.92 (d, 1H, J = 2.0 Hz), 7.05 (d, 1H, J = 9.0 Hz), 7.16 (d, 2H, J = 8.5Hz), 8.06 (d, 1H, J = 2.0 Hz), 8.07 (d, 1H, J = 9.0 Hz), 12.11 (s, 1H); FT-IR (film) v 3391, 2932, 1644, 1614, 1514, 1454, 1414, 1378, 1307, 1248, 1204 cm⁻¹; HR-ESI-FT-ICR-MS (m/z) [M + Na]⁺, calcd for C₃₅H₃₂NaO₁₀ 635.1888, found 635.1892.

Diol 20. To a solution of triol 18 (24.1 mg, 0.0332 mmol) in THF (1 mL) was added 1,1'-carbonyldiimidazole (10.8 mg, 0.066 mmol). The mixture was stirred for 15 h at room temperature, and 1,1'-carbonyldiimidazole (5.4 mg, 0.033 mmol) was further added. The resulting mixture was stirred for 24 h and concentrated. The residue was purified by flash column chromatography (silica gel, hexane/AcOEt = 3 to 2) to give the corresponding carbonate (19.4) mg, 0.0258 mmol, 78%) as a colorless syrup: $[\alpha]_D^{22.0} - 57.7^\circ$ (c 0.388, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.63 (q, 6H, J = 7.5 Hz), 0.93 (t, 9H, J = 7.5 Hz), 2.55 (d, 1H, J = 18 Hz), 2.66 (s, 3H), 2.84 (d, 1H, J = 18 Hz), 3.63 (s, 1H), 3.73 (s, 3H), 3.84 (s, 3H), 4.41 (d, 1H, J = 5.0 Hz), 4.55 (dd, 1H, J = 7.5, 5.5 Hz), 4.57-4.66 (m, 4H, H14), 5.96 (dd, 1H, J = 5.0, 2.0 Hz), 6.21 (d, 1H, J = 2.0 Hz), 6.73 (d, 2H, J = 8.5 Hz), 6.92 (d, 1H, J = 1.5Hz), 7.05 (d, 1H, J = 9.5 Hz), 7.16 (d, 2H, J = 8.5 Hz), 8.05 (d, 1H, J = 1.5 Hz), 8.08 (d, 1H, J = 9.5 Hz), 12.08 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) & 6.4, 7.2, 20.4, 29.6, 55.2, 55.4, 55.4, 63.6, 67.3, 73.1, 74.8, 80.8, 82.4, 85.1, 89.7, 90.4, 91.0, 94.2, 104.2, 105.2, 114.1, 115.9, 116.4, 123.3, 129.3, 129.5, 131.8, 133.3, 133.6, 134.2, 137.4, 153.9, 159.7, 159.7, 165.4, 172.0; FT-IR (film) v 2955, 2876, 1821, 1644, 1615, 1514, 1464, 1412, 1377, 1344, 1306, 1248, 1205 cm⁻¹; HR-ESI-FT-ICR-MS (m/z) [M+Na]⁺, calcd for C₄₂H₄₄-NaO₁₁Si 775.2545, found 775.2546.

To a solution of the above carbonate (19.4 mg, 0.0258 mmol) in THF (1 mL) was added TBAF (1.0 M solution in THF, 38.7 μ L, 0.0387 mmol) at 0 °C. The mixture was stirred for 40 min at 0 °C and concentrated. The residue was purified by flash column chromatography (silica gel, hexane/AcOEt = 1.5 to 0.5 followed by AcOEt/MeOH = 20) to give diol 20 (13.9 mg, 0.0218 mmol, 84%) as a colorless solid: $[\alpha]_D^{22.0} - 119^\circ$ (c 0.560, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 2.64 (s, 3H), 2.71 (d, 1H, J = 18 Hz), 2.78 (br s, 1H), 2.87 (d, 1H, J = 18 Hz), 3.65 (s, 3H), 3.71 (s, 1H), 3.72 (s, 3H), 4.41 (d, 1H, J = 5.0 Hz), 4.55–4.59 (m, 2H), 4.64 (d, 1H, J = 12 Hz), 4.66 (t, 1H, J = 11 Hz), 4.70 (d, 1H, J = 12 Hz), 6.02 (dd, 1H, J = 5.0, 2.0 Hz), 6.21 (d, 1H, J = 2.0Hz), 6.67 (d, 2H, J = 9.0 Hz), 6.89 (br s, 1H), 7.05 (d, 1H, J = 9.5 Hz), 7.16 (d, 2H, J = 9.0 Hz), 7.90 (br s, 1H), 8.07 (d, 1H, J = 9.5 Hz), 11.99 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 20.2, 28.4, 55.1, 55.2, 55.2, 63.3, 67.2, 73.2, 74.3, 80.2, 82.8, 83.6, 88.2, 88.3, 88.8, 94.6, 104.2, 104.2, 114.0, 116.1, 116.6, 123.2, 128.9, 129.6, 131.8, 132.3, 133.2, 134.2, 137.1, 153.6, 159.6, 159.6, 165.1, 171.8; FT-IR (film) v 3387, 2932, 1820, 1644, 1614, 1514, 1465, 1412, 1378, 1307, 1249, 1204 cm⁻¹; HR-ESI-FT-ICR-MS (m/z) [M + Na^{+} , calcd for $C_{36}H_{30}NaO_{11}$ 661.1680, found 661.1677.

TES Ether 21. To a solution of diol **20** (35.9 mg, 0.0562 mmol) in CH₂Cl₂ (2 mL) were added H₂O (100 μ L) and DDQ (38.3 mg, 0.169 mmol). The mixture was stirred for 10 h at room temperature and quenched with saturated NaHCO₃ solution and then saturated Na₂S₂O₃ solution. The resulting mixture was extracted with EtOAc $(3\times)$, and the combined organic layer was washed with brine and dried over anhydrous MgSO₄. After concentration, the residue was purified by flash column chromatography (silica gel, hexane/ AcOEt = 2 to 0.5) to give the corresponding triol (23.5 mg, 0.0453) mmol, 81%) as a colorless solid: $[\alpha]_D^{23.0} - 39.0^\circ$ (*c* 0.384, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 2.63 (s, 3H), 2.65 (d, 1H, J = 18Hz), 2.83 (br s, 1H), 2.97 (d, 1H, J = 18 Hz), 3.63 (br s, 1H), 3.71 (s, 1H), 3.90 (s, 3H), 4.51 (d, 1H, J = 4.5 Hz), 4.53 (dd, 1H, J =8.0, 5.5 Hz), 4.61 (dd, 1H, J = 9.0, 5.5 Hz), 4.68 (t, 1H, J = 8.5Hz), 5.73 (dd, 1H, J = 4.5, 2.0 Hz), 6.22 (d, 1H, J = 2.0 Hz), 6.91 (d, 1H, J = 2.0 Hz), 7.04 (d, 1H, J = 9.0 Hz), 7.98 (d, 1H, J = 2.0Hz), 8.08 (d, 1H, J = 9.0 Hz), 11.83 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 20.2, 28.2, 55.3, 55.4, 63.4, 67.3, 74.5, 83.4, 83.5, 84.0, 84.3, 88.3, 88.9, 94.7, 103.7, 104.1, 116.1, 116.6, 123.2, 130.9, 132.5, 133.7, 134.1, 137.4, 153.7, 159.8, 165.4, 173.4; FT-IR (film) v 3461, 2930, 1818, 1790, 1644, 1615, 1454, 1413, 1379, 1310, 1265, 1206 cm⁻¹; HR-ESI-FT-ICR-MS (m/z) [M + Na]⁺, calcd for C₂₈H₂₂NaO₁₀ 541.1105, found 541.1107.

To a solution of the above triol (19.2 mg, 0.0370 mmol) in CH₂-Cl₂ (2 mL) were added 2,6-lutidine (129 μ L, 1.11 mmol) and (TES)-

OTf (66.9 μ L, 0.296 mmol) at -90 °C. The mixture was stirred for 15 min at -90 °C and quenched with saturated NaHCO3 solution. The resulting mixture was extracted with AcOEt $(2\times)$, and the combined organic layer was washed with brine and dried over anhydrous Na₂SO₄. After concentration, the residue was purified by flash column chromatography (silica gel, hexanehexane/AcOEt = 4 to 3 to 2) to give bis(TES ether) 21 (19.5 mg, 0.0261 mmol, 71%) as a pale yellow syrup: $[\alpha]_D^{21.5} - 17.7^\circ$ (c 0.390, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.56 (q, 6H, J = 8.0 Hz), 0.81 (q, 6H, J = 8.0 Hz), 0.85 (t, 9H, J = 8.0 Hz), 1.00 $(t, 9H, J = 8.0 \text{ Hz}), 2.58 \text{ (d, 1H, } J = 18 \text{ Hz}), 2.61 \text{ (s, 3H)}, 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, 3H)}, 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{ Hz}), 2.76 \text{ (d, } J = 18 \text{ Hz}), 2.61 \text{ (s, } 3 \text{$ 1H, J = 18 Hz), 3.69 (s, 1H), 3.84 (s, 3H), 4.45 (d, 1H, J = 5.0Hz), 4.57 (dd, 1H, J = 7.3, 5.0 Hz), 4.60 (dd, 1H, J = 7.3, 5.0 Hz), 4.67 (t, 1H, J = 7.5 Hz), 5.57 (dd, 1H, J = 5.0, 2.0 Hz), 6.36 (d, 1H, J = 2.0 Hz), 6.88 (d, 1H, J = 2.0 Hz), 6.90 (d, 1H, J = 2.0Hz), 6.94 (d, 1H, J = 9.0 Hz), 7.85 (d, 1H, J = 9.0 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 4.8, 5.5, 6.7, 6.8, 19.7, 28.1, 55.0, 55.2, 63.3, 67.1, 74.3, 82.6, 82.7, 83.3, 88.7, 89.4, 94.1, 100.8, 116.9, 117.8, 118.5, 123.6, 127.7, 131.2, 132.9, 133.3, 136.5, 152.0, 153.7, 158.7, 168.6; FT-IR (film) v 2956, 2877, 1822, 1730, 1621, 1512, 1465, 1410, 1338, 1267, 1205 cm⁻¹; HR-ESI-FT-ICR-MS (m/z) [M + $Na]^+$, calcd for $C_{40}H_{50}NaO_{10}Si_2$ 769.2835, found 769.2839.

Neocarzinostatin Chromophore Aglycon (2). To a solution of alcohol **21** (3.9 mg, 0.0052 mmol) in CH₂Cl₂ (1.2 mL) was added Martin sulfurane dehydrating reagent (23.0 mg, 0.0340 mmol). The mixture was stirred for 10 min at room temperature and concentrated to 0.1 mL. A solution of trifluoroacetic acid–THF–water (1:10:5) was added, and the mixture was stirred for 35 min at 0 °C. Et₂O (10 mL) and saturated NaHCO₃ solution (5 mL) were added, and the resulting mixture was stirred for 1 h at 0 °C. The aqueous phase was extracted with Et₂O–AcOEt (2×), and the combined organic layer was dried over anhydrous MgSO₄. After filtration, the solution was concentrated to 0.3 mL and exposed to flash column chromatography (silica gel, hexane/AcOEt = 1.5 at 0 °C). The fractions containing aglycon were pooled, concentrated to 0.2 mL, and then

subjected to HPLC (Mightysil Si 60, 250-10 (5 µm), flow rate 4 mL/min, eluent hexane/AcOEt = 1, UV detection 254 nm). The fractions including pure aglycon 2 (retention time 10 min 15 s) were concentrated to 0.2 mL, diluted with MeOH (3 mL), and concentrated to 0.2 mL. This process was repeated, and the concentrate was taken up in MeOH (5 mL). A 0.5 mL solution was diluted to 10 mL with MeOH. UV absorbance was determined to quantitate 2 ($A_{302} = 0.51$, $\epsilon_{302} = 7040$ M⁻¹ cm⁻¹,^{16c} 2.3 mg, 0.0032 mmol, 61% in two steps from 21): ¹H NMR (400 MHz, CD₃CN) δ 2.60 (s, 3H), 3.83 (s, 3H), 4.06 (d, 1H, J = 1.6 Hz), 4.24 (brd, 1H, J = 5.2 Hz), 4.39 (dd, 1H, J = 9.2, 5.2 Hz), 4.62 (t, 1H, J = 8.8 Hz), 4.91 (dd, 1H, J = 8.4, 5.6 Hz), 4.89–4.93 (m, 1H), 5.66 (br s, 1H), 6.01 (t, 1H, J = 3.0 Hz), 6.67 (br s, 1H), 6.92 (br s, 1H), 7.04 (d, 1H, J = 9.2 Hz), 7.95 (br s, 1H), 8.13 (d, 1H, J = 9.2 Hz), 11.59 (s, 1H); FT-IR (film) v 3677, 3068, 2926, 1820, 1739, 1613, 1465, 1380 cm⁻¹; HR-ESI-FT-ICR-MS (m/z) [M + Na]⁺, calcd for C₂₈H₂₀NaO₉ 523.1000, found 523.1002; CD (MeOH) λ_{ext} 330.4 nm ($\Delta \epsilon$ +1.06), 314.7 (0.0), 294.1 (-1.92), 289.8 (-1.86), 264.1 (-5.84), 243.9 (0.0).

Acknowledgment. This work was supported by a Grantin-Aid for Scientific Research (Wakate B) from the Japan Society for the Promotion of Science (JSPS). We are grateful to Mr. Hiroyuki Monma, Research and Analytical Center for Giant Molecules, Tohoku University, for measurement of ESI-FT-ICR mass spectrometry, and to Prof. Masahiro Toyota, Osaka Prefecture University, for his generous support.

Supporting Information Available: ¹H and ¹³C NMR spectra of compounds **4**–**22**, and ¹H NMR, UV, CD, and mass spectra of aglycon **2**. This material is available free of charge via the Internet at http://pubs.acs.org.

JO052031O